

Investigation of a Chemical Model for the Methylation of Deoxyuridine Monophosphate by Thymidylate Synthetase

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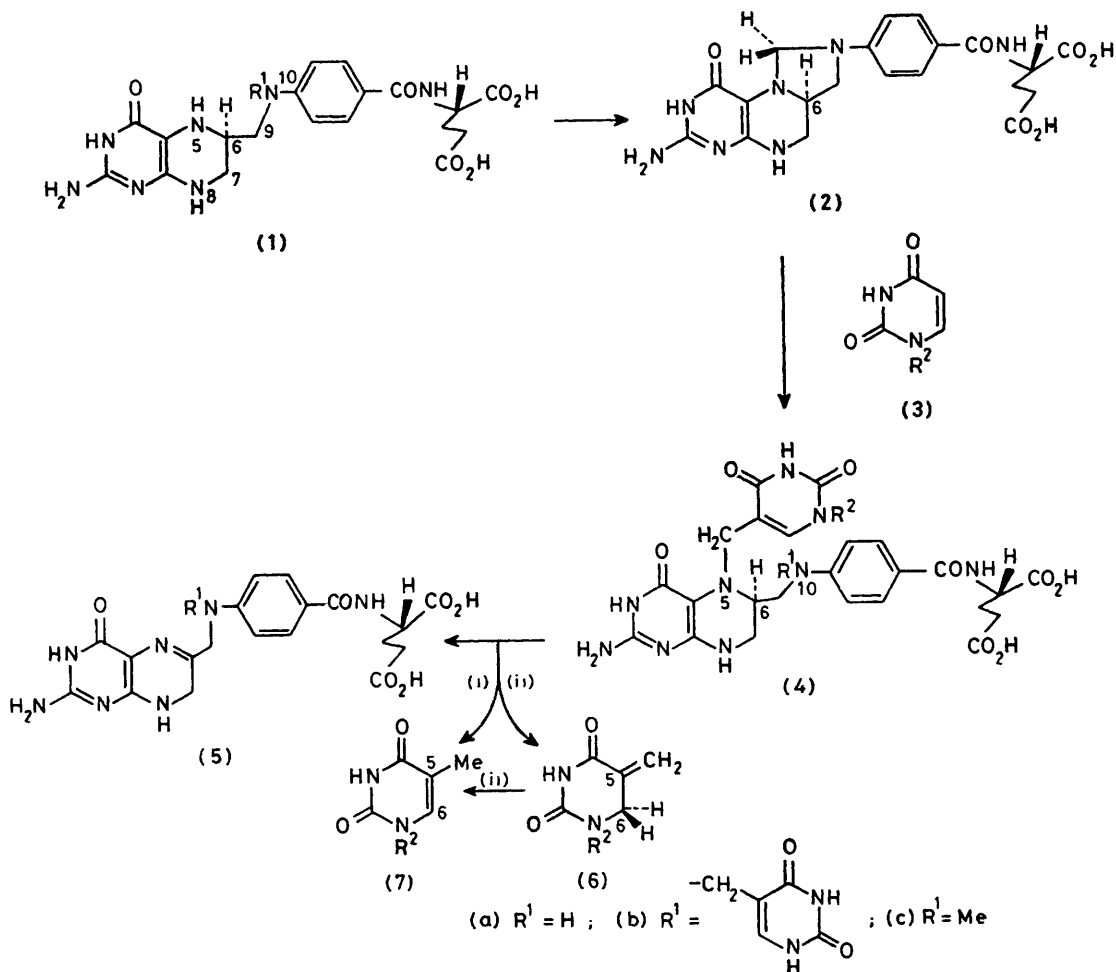
Summary The compound (**4a**, $R^2 = H$), long regarded as a putative intermediate in a chemical model for thymine biosynthesis, has been reformulated as the *bis*-adduct (**4b**; $R^2 = H$), a chemical model for the action of thymidylate synthetase has been achieved and the result is discussed

THYMIDYLATE SYNTHETASE, the enzyme responsible for methylation of the RNA base uracil to the DNA base thymine, is an important target enzyme in cancer chemotherapy¹. This methylation is mediated by the coenzyme 5,6,7,8-tetrahydrofolic acid (**1**, $R^1 = H$) and the one-carbon transfer involves 5,10-methylene-5,6,7,8-tetrahydrofolic acid (**2**) rather than 5-methyl-5,6,7,8-tetrahydrofolic acid which

is the normal substrate for one-carbon transfers at this oxidation level.² The reduction concomitant with the transfer is achieved by 'oxidation' of the cofactor to 7,8-dihydrofolic acid (5; $R^1 = H$), and it is known that the hydrogen at C-6 of 5,6,7,8-tetrahydrofolate (1; $R^1 = H$) is transferred to the methyl group of thymine (7; $R^2 = H$) in this process.³

shift might then complete the sequence [Scheme (ii)]. We now report work on the thermal conversion of a compound of type (4) into thymine which would be a chemical model for this proposed biochemical process.

Since Gupta and Huennkens⁶ had reported a synthesis of (4a; $R^2 = H$) by alkylation of 5,6,7,8-tetrahydrofolic acid with two moles of 5-chloromethyluracil, we reduced 7,8-



SCHEME

One suggestion as to how this sequence might be carried out chemically is outlined in the Scheme. A uracil derivative (3) might undergo Mannich reaction with the cofactor (2) to yield the product (4). This intermediate might then rearrange to the thymine derivative (7) [Scheme (i)] and 7,8-dihydrofolic acid (5). The intermediacy of compounds of type (4) was first suggested by Friedkin⁴ and a nominal 1,3-hydride shift from C-6 of (4) to the methyl of thymine would be required to complete the process. Since a *concerted* 1,3-hydride shift is not allowed,⁵ it is appealing to consider the possibility that the intermediate (4) might undergo an allowed concerted *retro-ene* reaction to yield 7,8-dihydrofolic acid (5) and the intermediate (6) in which the hydrogen at C-6 in (4) would be delivered to C-6 of the pyrimidine (6) [Scheme (ii)]. A stereospecific enzyme-catalysed proton

dihydrofolic acid (5)⁷ with $NaBH_4$, took the pH to 0–1 with conc. HCl and alkylated the product at pH 7.5 with two moles of 5-chloromethyluracil. The product had u.v. spectra at acid, neutral, and basic pH which were identical in every way with those reported for (4a; $R^2 = H$).⁶ The compound ran as a single quenching spot on paper chromatography and analysed as $C_{24}H_{27}N_9O_8 \cdot 4H_2O$. Coincidentally the analytical figures also fitted a *bis*-adduct, $C_{29}H_{31}N_{11}O_{10} \cdot 4H_2O$. The 1H -n.m.r. spectrum^{8a} of the compound in 1 M NaOD- D_2O showed, in addition to the typical tetrahydrofolic acid resonances, a one-proton singlet at δ 7.05, a one-proton doublet (J 1.8 Hz) at δ 7.79, a two-proton AB system centred at δ ca. 3.6 and a further two protons overlapping with the α -CH of the glutamate residue at δ ca. 4.2. It would appear, therefore, that the product was the *bis*-adduct

(**4b**, $R^2 = H$) and this was confirmed by a field desorption mass spectrum^{8b} which showed a major ion at m/e 694 ($C_{29}H_{31}N_{11}O_{10}$ requires 693.64)

In view of the similarity of synthesis and the identity of the spectra of our product with those of the compound previously assigned structure (**4a**, $R^2 = H$),⁶ we repeated the further alkylation of this compound.⁶ This reaction had afforded a product claimed⁶ to be (**4b**, $R^2 = H$). The 1H -n m r spectrum^{8a} of the crude product indicated that peralkylation had occurred.

Although we had obtained the *bis*-adduct (**4b**, $R^2 = H$), we resolved to investigate its pyrolysis. 1H -N m r spectra indicated that, at most, a negligible amount of thymine was present in the compound, but when it was heated to 255 °C *in vacuo*, thymine sublimed in 39% yield. The [6- 2H_1]-analogue of (**4b**, $R^2 = H$) was prepared by substituting NaB^2H_4 for $NaBH_4$ in the synthesis and on pyrolysis this gave a 46% yield of thymine which was shown to be 29.1% monodeuteriated by mass spectroscopy. 2H -N m r spectroscopy^{8c} indicated that the isotope was located entirely on the methyl group of thymine (δ -2.93 from 2H_2O).

These results, while interesting, were confused by the presence of the second thymine residue at N-10. It was necessary therefore to synthesise a compound which contained only the N-5 thymine unit. To this end methotrexate was hydrolysed to 10-methylfolic acid with 1 M NaOH.⁹ The protected compound was now reduced with dithionite to

7,8-dihydro-10-methylfolic acid (**5**, $R^1 = Me$)[†]. Further reduction with $NaBH_4$ yielded (**1**, $R^1 = Me$) and this was alkylated *in situ* to yield the product (**4c**, $R^2 = H$)[‡]. The [6- 2H_1]-analogue[‡] of (**4c**, $R^2 = H$) was prepared by substituting NaB^2H_4 for $NaBH_4$ in the synthesis and both compounds were shown to contain, at most, a negligible amount of thymine by 1H -n m r spectroscopy.^{8a} Pyrolysis of both compounds gave thymine. The [6- 2H_1]-analogue gave a 47% yield of thymine which was shown to be 25.3% monodeuteriated by mass spectroscopy. 2H -N m r spectroscopy^{8c} again showed that all of the label was in the methyl group of thymine.

It is evident that the *retro*-ene mechanism for the biochemical reaction is ruled out by the isotopic experiments, since C-6 of thymine would be labelled in this pathway. The labelling pattern is in fact the same as is found in nature and a radical cage mechanism would seem to account best for the high discrimination for the hydrogen at C-6 of (**4**) over all of the other hydrogens present. A nominal 1,3-hydride shift of this type has been observed in the pyrolysis of dihydroquinoline adducts¹⁰ and a radical cage mechanism may be implied here also. An Arrhenius calculation suggests $t_{1/2}$ for radical formation to be *ca.* 2 s at 250 °C.

We thank the S R C for financial support and for a studentship (to P A C).

(Received, 28th March 1980, Com 332)

† A satisfactory 1H -n m r spectrum was obtained for this compound.

‡ This compound had satisfactory analytical and spectroscopic properties.

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⁴ See M Friedkin, *Adv Enzymol Relat Areas Mol Biol*, 1973, **38**, 235

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⁷ R L Blakley, *Nature* 1960, **188**, 231

⁸ We thank (a) Dr John C Lindon Wellcome Research Laboratories Beckenham, for these spectra and for helpful discussions on their interpretation, (b) Dr David A Brent, Burroughs Wellcome Co, U S A, for field desorption mass spectra, (c) Mr David R Stanley, A R C Unit of Nitrogen Fixation, Sussex, for 2H -n m r spectra

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